

Exploring the scientific and technological potential of medicinal plants for anthelmintic therapies

Explorando o potencial científico e tecnológico de plantas medicinais para terapias anti-helmínticas

Maria Crisnanda Almeida Marques¹, Rusbene Bruno Fonseca de Carvalho², Solange Sousa Santos³, Francisco Mayron de Sousa e Silva⁴, Chistiane Mendes Feitosa⁵, Livio Cesar Cunha Nunes⁶

ABSTRACT

Parasitic infections are prevalent in tropical and underdeveloped regions, where poor sanitary conditions allow parasites to thrive. The common practice of administering anthelmintic drugs without prior diagnosis has led to increasing parasite resistance. An alternative approach involves the use of medicinal plants, traditionally employed in folk medicine for their anthelmintic properties. This article aims to explore the potential of medicinal plants for anthelmintic therapies, including an analysis of relevant scientific research and patents. Comprehensive searches were conducted in scientific databases (Science Direct, PubMed) and patent databases (WIPO, INPI, USPTO, EPO). A total of 32 scientific articles meeting our criteria were evaluated, focusing on plant species used, microorganisms tested, extraction methods, and outcomes. Results show that various plant parts are employed against parasites affecting both human and animal health, with many studies isolating bioactive compounds for testing. Additionally, 495 patents were reviewed, with 6 meeting the eligibility criteria—all filed by India. Despite the promising potential of medicinal plants in anthelmintic therapies, the number of patent filings remains low. However, scientific interest and advancements in the use of plant species for treating parasitic diseases are evident and show significant promise for future applications.

Keywords: Parasitic diseases. Treatment. Phytotherapy.

RESUMO

Infecções parasitárias são prevalentes em regiões tropicais e subdesenvolvidas, onde condições sanitárias precárias favorecem a proliferação de parasitas. A prática comum de administração de fármacos anti-helmínticos sem diagnóstico prévio tem contribuído para o aumento da resistência parasitária. Uma abordagem alternativa envolve o uso de plantas medicinais, tradicionalmente empregadas na medicina popular por suas propriedades anti-helmínticas. Este artigo tem como objetivo explorar o potencial das plantas medicinais em terapias anti-helmínticas, incluindo uma análise de pesquisas científicas e patentes relevantes. Foram realizadas buscas abrangentes em bases de dados científicas (Science Direct, PubMed) e de patentes (WIPO, INPI, USPTO, EPO). Um total de 32 artigos científicos que atenderam aos critérios de inclusão foi avaliado, com foco nas espécies vegetais utilizadas, nos microrganismos testados, nos métodos de extração e nos resultados obtidos. Os achados indicam que diversas partes de plantas são empregadas no combate a parasitas que afetam a saúde humana e animal, sendo que muitos estudos buscaram isolar compostos bioativos para testes específicos. Além disso, 495 patentes foram analisadas, das quais 6 atenderam aos critérios de elegibilidade—todas depositadas pela Índia. Apesar do potencial promissor das plantas medicinais em terapias anti-helmínticas, o número de depósitos de patentes permanece reduzido. Contudo, observa-se crescente interesse científico e avanços no uso de espécies vegetais para o tratamento de doenças parasitárias, evidenciando perspectivas significativas para aplicações futuras.

Palavras-chave: Doenças parasitárias. Tratamento. Fitoterapia.

¹ Master's degree student, Graduate Program in Pharmaceutical Sciences, Federal University of Piauí.
ORCID: 0000-0002-0554-001X
Email: mariacrisnanda@ufpi.edu.br

² Researcher, Federal University of Piauí.
ORCID: 0000-0002-5993-1729
Email: rusbenecarvalho@gmail.com

³ PhD student, Graduate Program in Pharmaceutical Sciences, Federal University of Piauí.
ORCID: 0000-0003-0963-5312
Email: solange.santos@gmail.com

⁴ PhD student, Graduate Program in Pharmaceutical Sciences, Federal University of Piauí.
ORCID: 0000-0002-3916-880X
Email: mayrondcf@gmail.com

⁵ Researcher, Graduate Program in Pharmaceutical Sciences, Federal University of Piauí.
ORCID: 0000-0001-8013-1761
Email: chistiane@ufpi.edu.br

⁶ Researcher, Graduate Program in Pharmaceutical Sciences, Federal University of Piauí.
ORCID: 0000-0002-1178-7940
Email: liviocsar@hotmail.com

1. INTRODUCTION

Helminthiasis are prevalent in developing tropical regions, particularly in areas with precarious sanitation, affecting over one-third of the global population¹. Transmission occurs via soil, with *Ascaris lumbricoides*, *Trichuris trichiura*, hookworms (*Ancylostoma duodenale*, *Necator americanus*), *Strongyloides stercoralis*, and *Enterobius vermicularis* being the most common². These parasites infect the gastrointestinal tract through ingestion of eggs or larvae, with an estimated 300 million individuals severely affected—many of them schoolchildren³—who are also more susceptible to co-infections and higher mortality risk⁴.

In high-prevalence regions, mass drug administration is recommended, mainly for school-age children⁵, using albendazole or mebendazole without prior diagnosis⁶. However, frequent and improper use has promoted resistance and the selection of genes linked to autoimmunity, reinforcing the need to identify new anthelmintic substances^{7,8}. Medicinal plants emerge as a viable alternative, given their traditional antiparasitic use and potential for isolating active compounds such as tannins, flavonoids, terpenes, and alkaloids⁹. Metabolites including cinnamic acid, carvacrol, thymol, and gallic acid have demonstrated proven anthelmintic activity¹⁰. Species such as *Punica granatum*, *Ginkgo biloba*, *Thymus vulgaris*, and *Carica papaya*—tested via extracts, essential oils, and latex—exhibited defined mechanisms of action¹⁰. Brazil's biodiversity offers a rich source for such studies, as many native species with traditional anthelmintic use are distributed across its six biomes⁹. Nevertheless, research remains limited, justifying this study's aim to survey scientific and technological developments on plant species with anthelmintic potential.

2. MATERIALS AND METHODS

This quantitative, descriptive study employed scientific and technological prospecting on medicinal plant-based anthelmintic therapies, following Silva et al.¹¹. Literature searches in Science Direct and PubMed applied the PRISMA protocol¹², using the MeSH terms “Medicinal Plants” AND “Anthelmintics” in titles, abstracts, and keywords. Publications (2014–2024) in English were included; reviews, conference papers, theses, editorials, non-

original reports, and duplicates were excluded. Titles, abstracts, and full texts were screened for data extraction. Data were managed in StArt software (LaPES/UFSCar).

Patent searches were conducted in INPI, WIPO (Patentscope), EPO, and USPTO databases, using “Medicinal Plants” AND “Anthelmintics” (Portuguese for INPI, English for others). Inclusion criteria: classification under A61K, publication within the last 10 years, and relevance to anthelmintic activity against human parasites or experimental models. Screening involved titles, abstracts, and full-text eligibility confirmation. Extracted data included title, author, year, plant species, form tested, concentration, target parasite, and country of origin.

3. RESULTS

A review of the studies in Table 1 confirms the therapeutic potential of medicinal plants for helminth control. Various species and their phytochemicals have shown efficacy against *Haemonchus placei*, *Haemonchus contortus*, and *Gyrodactylus kobayashii*. Standardized extraction methods and diverse *in vitro* and *in vivo* assays enabled the identification of bioactive molecules with significant anthelmintic activity. Prominent results were observed for *Sarcocephalus latifolius*, *Kyllinga odorata*, *Chenopodium album*, and *Jasminum grandiflorum* L. subsp. *floribundum*, with compounds such as plumbagin, flavonoids, essential oils, and rutin emerging as promising candidates. These findings reinforce the value of ethnopharmacological approaches and the need for continued bioprospecting of botanical resources.

Table 1 - Exploring the Potential of Medicinal Plants: An Anti-helminthic Approach.

REF.	PART OF THE PLANT	OBJECTIVE	MICROORG.	METHOD	BIOLOGICAL TESTS	OUTCOME
13	<i>Lavandula angustifolia</i> e <i>Quercus infectoria</i> (essential oils)	Investigating anthelmintic activity against the parasite <i>Marshallagia marshalli</i>	Parasite <i>Marshallagia marshalli</i>	Egg hatching tests, inhibition of adult and larval motility, DNA damage, evaluation of oxidative/nitrosative stress biomarkers	Egg hatching tests, inhibition of adult and larval motility, assessment of DNA damage and oxidative/nitrosative stress biomarkers	Essential oils showed significant activity against the parasite, with effects on egg hatching, motility and DNA damage
14	<i>Sarcocephalus latifolius</i> (leaves)	Identify and characterize anthelmintic compounds.	<i>Haemonchus placei</i> (verme adulto)	Leaf extraction and chromatographic separations.	Anthelmintic activity and cytotoxicity against mammalian cells	Bioactive compounds with anthelmintic activity identified

15	<i>M. inermis</i> (Henna) (leaves)	Identify anthelmintic substances	<i>Haemonchus contortus</i>	Phytochemical screening, UPLC-QToF-ESI-MS analysis	In vitro tests on <i>H. contortus</i> , toxicity test on <i>Artemia salina</i> larvae	Extracts of the leaves showed anthelmintic activity in vitro, with no significant toxicity
16	<i>Morinda lucida</i> <i>Benth</i> (Rubiaceae)	Identify anthelmintic compounds	<i>H. contortus</i> and <i>C. elegans</i>	Extract fractionation, HPLC-HRMS/MS analysis, cytotoxicity test	Anthelmintic activity tests on young adults of <i>C. elegans</i> , cytotoxicity test on WI38 cells	Active fractions identified, potential compounds suggested, need for further studies to confirm
17	<i>Euphorbia forskalii</i> (whole plants)	Evaluate in vitro anthelmintic activity against eggs, infesting larvae (L3) and adult worms of <i>Haemonchus contortus</i> .	<i>Haemonchus contortus</i>	Preparation of aqueous extracts from dried and grounded plants.	Tests for adult worm mortality, inhibition of egg hatching, inhibition of larval migration	Extracts of <i>E. forskalii</i> showed anthelmintic activity in vitro against adult worms of <i>H. contortus</i> , and significant inhibition of egg hatching and larval migration
18	<i>Ceiba pentandra</i> (stem bark)	Evaluate anthelmintic activity against <i>Haemonchus contortus</i>	Eggs, infective larvae (L3) and adult worms of <i>Haemonchus contortus</i>	In vitro tests (egg hatching test, inhibition of larval migration, inhibition of adult worm motility)	In vitro tests for different stages of parasites	<i>Ceiba pentandra</i> stem bark showed anthelmintic activity against <i>Haemonchus contortus</i>
19	<i>P. umbellata</i> (root extracts) and compound 4-nerolidylcatechol (4-NC)	Investigate anti-schistosomal effects against <i>Schistosoma mansoni</i>	Adult worms of <i>Schistosoma mansoni</i>	<i>Ex vivo</i> and <i>in vivo</i> tests and biochemical analysis	Tests on <i>Schistosoma mansoni</i> adults, murine models of schistosomiasis	Extracts and 4-NC showed significant anti-schistosomal activity, need for further studies
20	<i>Rumex crispus</i> L. (leaves)	Evaluate anthelmintic potency and identify bioactive compounds	<i>Caenorhabditis elegans</i>	Testing of solvent extracts for up to 72 hours; chemical analysis of compounds	Lethality tests and microscopic analysis	Confirmed anthelmintic potential; presence of various bioactive compounds
21	<i>Blumea lacera</i> (aerial parts)	Evaluate medicinal properties, including analgesic, anti-inflammatory, antidiarrheal, diuretic activity and acute toxicity.	<i>Paramphistomum cervi</i>	Extraction with 96% ethanol, pharmacological tests in mice.	Analgesic, anti-inflammatory, antidiarrheal, diuretic, acute toxicity, antioxidant, phytochemical, anthelmintic, cytotoxic tests	<i>B. lacera</i> has shown potential for treating various diseases, but has also shown toxicity in tests, suggesting the need to purify bioactive compounds
22	<i>Piper aduncum</i> (ethanolic extract)	Evaluate monogenetic control in <i>Colossoma macropomum</i>	<i>Colossoma macropomum</i> monogenetics	In vitro and in vivo tests; determination of CL50-24h	Physiological tests on treated fish	Efficacy of up to 99.2%; identification of dilapiol as the active component

23	<i>Spondias mombin</i> (leaves)	L.	Isolate and characterize anthelmintic compounds, establish bioactivity and safety profile.	<i>Fasciola hepatica</i>	Chromatographic fractionation, characterization by spectroscopy and LC-ESI-MS.	Screening assays in hepatic, cytotoxicity in MDBK cells	F.	Compound MC010 showed moderate anthelmintic activity against adult <i>Fasciola hepatica</i> worms, but with moderate toxicity in MDBK cells
24	<i>Cassia sieberiana</i> (root bark)		Validate traditional application as a vermifuge, analyze metabolic composition.	<i>Haemonchus contortus</i>	Extraction with 70% ethanol, chemical analysis, larval migration inhibition tests.	Larval migration inhibition tests		The ethyl acetate extract proved to be the most active, with a possible synergy between flavonoids and proanthocyanidins
25	Plumbagina, Timoquinona, Osthole, Psoraleno		Investigating the anthelmintic efficacy of phytochemical compounds.	<i>Gyrodactylus kobayashii</i> (monogenetic parasite)	Exposure of fish to different compounds and in vitro and in vivo tests.	Efficacy of the compounds against the monogenean parasite		Plumbagin was the most effective compound against <i>G. kobayashii</i>
26	Thymoquinone (TQ) of <i>Nigella sativa</i> seeds		Investigate the anthelmintic effect of TQ on adult worms of <i>Gigantocotyle explanatum</i>	<i>Gigantocotyle explanatum</i> adult worms	Treatment with different concentrations of TQ; analysis of ultrastructure, motility, antioxidant/detoxifying enzymes and DNA profiles	Electron microscopy, enzyme activity tests, DNA analysis		TQ inhibited glutathione metabolism, imbalanced the redox system and inhibited enzymes, reflecting anthelmintic potential, but further studies are needed
27	<i>Jasminum grandiflorum</i> L. subsp. <i>Floribundum</i> (aerial parts)		Fractionation and identification of anthelmintic compounds.	Various microorganisms (larvae, adults and cysts)	Extraction of the aerial parts and testing for anthelmintic activity.	Evaluation of the compounds on different microorganisms.		Flavonoids and secoiridoid glycosides have shown anthelmintic activity.
28	<i>Spondias mombin</i> L. (Anacardiaceae)		Isolating and characterizing anthelmintic compounds	<i>Haemonchus placei</i>	Bioassay-guided chromatographic fractionation, spectroscopy, LC-ESI-MS analysis	Cytotoxicity screening in mammalian cells		Pheophorbide-a isolate showed anthelmintic activity and low toxicity to mammalian cells.
29	<i>Cinnamomum bejolghota</i> (leaves)		Investigate anthelmintic and antihypertensive activity in vitro	<i>Caenorhabditis elegans</i> (Adult nematodes)	Column chromatography, compound isolation	Activity tests against adult nematodes, egg hatching, larval development		Geranial compound showed the greatest activity against adult nematodes and egg hatching
30	<i>Bombax ceiba</i> , <i>Diospyros rhodocalyx</i> , <i>Vitex glabrata</i> , <i>Terminalia</i>		Examine the anthelmintic efficacy of crude plant extracts and rutin against	<i>Gastrothylax crumenifer</i>	Obtaining crude plant extracts, fractionation bioassay and	Tests of anthelmintic activity of		N-butanol extract of <i>Terminalia catappa</i> leaves and rutin

	<i>catappa</i> and <i>Cassia alata</i>	<i>Gastrothylax</i> <i>crumenifer</i>		identification of rutin	extracts and rutin	showed greater anthelmintic activity against <i>Gastrothylax</i> <i>crumenifer</i>
31	<i>Cuscuta reflexa</i> (extracts)	Evaluate anti- diarrheal activity in pigeons	Causes diarrhea pigeons	of intramuscular administration of extracts; induction of diarrhea in pigeons	Tests to assess frequency and onset of diarrhea	Significant anti- diarrheal activity observed
32	<i>Tetrapleura</i> <i>tetraptera</i> (stem bark and fruits)	Investigation of the antitrypanosomal activity and anthelmintic activity	Blood form of <i>Trypanosoma</i> <i>brucei</i> ; <i>earthworms</i> <i>Pheretima</i> <i>posthuma</i>	In vitro antitrypanosomal and anthelmintic tests	In vitro antitrypanosomal and anthelmintic tests	Extracts showed antitrypanosomal and anthelmintic activity
33	<i>Ptaeroxylon</i> <i>obliquum</i> (leaves)	Determine anthelmintic, antifungal and antimycobacteri- al activities	Eggs and larvae of <i>Haemonchus</i> <i>contortus</i> ; fungal and bacterial microorganisms	In vitro tests for different activities and structural analysis	In vitro tests for different activities	Extract and obliquumol showed anthelmintic and antifungal activity
34	<i>Ziziphus</i> <i>mucronata</i> (unspecified part)	Provide an overview of pharmacological properties and biological activity	Various pathogens, including <i>Mycobacterium</i> <i>tuberculosis</i> , <i>Staphylococcus</i> <i>aureus</i> , etc.	Literature review	In vitro tests for various biological activities	<i>Ziziphus</i> <i>mucronata</i> has shown potential antimicrobial, antioxidant, anti- inflammatory and anthelmintic activity, among others.
35	<i>Acacia</i> <i>farnesiana</i> (pods)	Identify anthelmintic compounds against <i>H.</i> <i>contortus</i>	Eggs and larvae of <i>H. contortus</i>	Bioguided chemical fractionation of the pods; testing for anthelmintic activity	Tests on eggs and larvae and analysis of compounds	EtOAc-F fraction showed significant ovicidal and larvicidal activity
36	<i>Curcuma</i> <i>zedoaria</i>	To investigate anti-helminthic activities against <i>Gyrodactylus</i> <i>kobayashii</i>	<i>Gyrodactylus</i> <i>kobayashii</i>	Testing methanolic extracts and in vitro and in vivo studies	In vitro and in vivo tests; microscopic analysis	Extract and curdione compound showed anthelmintic activity
37	<i>Salvadora</i> <i>persica</i> L. (Root extracts)	Investigate the anthelmintic, anticoccidial and antioxidant activity of the extracts	<i>Allolobophora</i> <i>caliginosa</i> (anti- helminthic and <i>Eimeria papillata</i> (anticoccidial)	In vivo and in vitro tests of the extracts; biochemical analysis	Paralysis tests, oocyst count, biochemical analysis	Extracts have shown anthelmintic, anticoccidial and antioxidant activity, further studies needed
38	<i>Allium sativum</i> (garlic)	Determine in vivo efficacy in horses infected with strongyloides	Intestinal strongyles in horses	Field trial on mares and fecal egg reduction test	Fecal egg count reduction tests	Garlic has not shown significant efficacy
39	<i>Andrographis</i> <i>paniculata</i> (leaves)	Evaluate ovicidal and larvicidal effects against <i>Ancylostoma</i> <i>duodenale</i>	<i>Ancylostoma</i> <i>duodenale</i>	Tests of extracts against eggs and larvae and identification of diterpenoid compounds	Ovicidal and larvicidal activity tests	Extracts showed significant ovicidal and larvicidal activity
40	<i>Kniphofia foliosa</i> Hochst.	Describe activity against various microorganisms	<i>Plasmodium</i> <i>falciparum</i> , <i>elegans</i> , C. A.	Cytotoxicity and anti-parasitic activity tests	Cytotoxicity and anti-parasitic activity tests	Knipholone has shown activity against

			<i>fischeri</i> , <i>tuberculosis</i> , 1	<i>M.</i> HIV-				<i>Plasmodium</i> <i>falciparum</i>
41	<i>Baccharis conferta</i> (aerial parts)	Isolar ovidal composts against <i>Haemonchus contortus</i>	<i>Haemonchus contortus</i> eggs		Extraction and fractionation; ovidal activity tests	Ovidal activity tests	Fractions and isolated compounds inhibited egg hatching	

Source: Prepared by the authors.

From the technological innovation perspective, database searches retrieved 495 patents, of which 14 met the relevance criteria. After excluding patents without methodological clarity, outdated documents, and those for exclusively veterinary use, six remained (Table 2). All originated from India and were filed between 2010 and 2022, reflecting the country's significant helminthiasis burden—particularly *Ancylostoma duodenale*—and strategic investment in phytotherapeutic innovation. In the face of rising global drug resistance and the impact of helminthiasis on child health and quality of life, these patents represent critical steps toward accessible and effective alternatives to conventional treatments.

Table 2 - Survey of patent information obtained from the different databases used in this study.

REF.	TITLE	FORMULATION	PARASITE	INNOVATION	COUNTRY
42	Plant protection preparation useful for the treatment of filaria, especially as a macrofilaricidal agent	Ethanollic and methanollic extracts of <i>Calotropis</i> sp. and isolated compounds were tested at concentrations of 125 µg/ml and 62.5 µg/m	<i>Brugia malayi</i>	At a concentration of 125 µg/ml, this ethanolic extract showed 100% microfilaricidal activity. At this concentration, the extract had an IC50 value of no more than 52.6 µg/ml. The selectivity index (SI) in relation to adult motility was not less than 64.8.	India
43	Microencapsulated herbal product with ovidal and larvicidal properties against <i>Ancylostoma duodenale</i> (hookworm)	Microencapsulated <i>Andrographis paniculata</i> extract, concentrations tested against <i>Ancylostoma duodenale</i> were 0.25 to 5mg	<i>Ancylostoma duodenale</i>	The microencapsulated product was able to reduce the parasite count, with a rate of 100% at a concentration of 5 mg, a value similar to and even higher than the positive control.	India
44	A new formulation of <i>Chenopodium album</i> for antelmintic activity	<i>Chenopodium album</i> plant extracts prepared with petroleum ether, ethyl acetate, methanol, hydroalcoholic and aqueous solvents tested at concentrations of 2-10mg/ml	<i>Eisenia foetida</i> (experimental model)	All extracts lead to paralysis of <i>Eisenia foetida</i> except the hydroalcoholic extract, the ethyl acetate extract was effective at all concentrations tested.	India
45	A new formulation of <i>Operculina</i>	Extract of the roots/rhizomes of	<i>Eisenia foetida</i>	All extracts (except the aqueous extract) lead to	India

	<i>turpethum</i> for antelmintic activity	<i>Operculina turpethum</i> prepared with petroleum ether, ethyl acetate, methanol, hydroalcoholic and aqueous solvents	(experimental model)	paralysis and earthworm fatality. The OTPE extract exhibits potential anthelmintic activity at a dose of 10 mg/ml, causing paralysis and earthworm fatality and is more potent than the PCT suspension. At a dose of 10 mg/ml, the PT and DT times for OTPE were recorded as (9.38 ± 1.82) and (54.93 ± 1.78) .	
46	<i>Cocos nucifera</i> (coconut) shell ash as an anthelmintic drug substance	<i>Cocos nucifera</i> ash was tested at concentrations of 2, 4, 8, 10 and 20 mg/ml	<i>Pheretima posthuma</i> and <i>Haemonchus contortus</i>	The ashes proved to be effective when comparing the time to death of the concentrations tested with that obtained with the standard, at the same concentration of 4mg/ml they showed a time to death of 20 ± 0.38 and 17 ± 0.36 for the control and test respectively.	India
47	Comparative in vitro anthelmintic activity of <i>Adhatoda vasica</i> bark and <i>Momordica charantia</i> against <i>Pheretima posthuma</i>	Ethanol extract of the bark of <i>Adhatoda vasica</i> and <i>Momordica Charantia</i> against <i>Pheretima Posthuma</i> at concentrations of 10, 20, 25 and 50 mg/ml in the in vitro test and 100, 150 and 200mg/ml in the in vivo test.	<i>Pheretima Posthuma</i> (experimental model)	The death time of <i>Pheretima Posthuma</i> was evaluated, and the percentage of diarrhea inhibition was 55.27%, 66.33% and 63.81% at body doses of 100, 150 and 200mg/Kg, respectively.	India

Source: Prepared by the authors.

4. DISCUSSION

The analyzed results confirm that medicinal plants harbor diverse bioactive compounds with potential efficacy against multiple parasites. However, variability in effects, coupled with the need for further studies on safety and dosage, underscores the complexity of their therapeutic use. The following discussion addresses anthelmintic effects, extraction methods, and safety–efficacy evaluation.

4.1 Infectious and Parasitic Diseases

Infectious parasitic diseases (IPDs), caused by helminths with varied transmission cycles, present significant clinical and epidemiological complexity and affect populations in socially and environmentally vulnerable areas⁴⁸. Although mortality from IPDs has declined in Brazil in recent decades, rates remain high compared to other countries⁴⁸. These diseases affect both humans and animals; in humans, examples include malaria, Chagas disease,

and ascariasis⁴⁸.

4.2 Medicinal Plants in the Treatment of Parasitic Diseases

Medicinal plants have been investigated for parasite control in human and veterinary contexts. They offer a wide range of chemical compounds that may provide therapeutic benefits but can also pose risks. Proper identification, conservation, preparation, and knowledge of adverse effects are essential⁴⁹. Some species, such as *Chenopodium ambrosioides* and *Mentha suaveolens*, are effective against enteroparasitoses⁵⁰, while green tea and mint have shown activity against *Entamoeba histolytica* and *Giardia*, partly by altering intestinal pH and inhibiting parasite proliferation⁵¹.

4.3 Bioactive Compounds in Medicinal Plants: Examples and Sources

The bioactive profile of medicinal plants varies with species, plant part, and dosage⁵²⁻⁵³. Multiple plant parts—leaves, roots, rhizomes, aerial structures, stem bark, and fruits—have been studied for anthelmintic activity^{10,54-56}. Leaves, due to high metabolic activity and sunlight exposure, often accumulate secondary metabolites⁵⁴, while roots and rhizomes may contain compounds that protect against soil pathogens. Aerial parts also harbor diverse metabolites due to environmental exposure^{10,54-56}.

The choice of plant part is influenced by traditional medicinal practices^{10,54}. Analysis of Table 1 shows leaves as the most investigated structures, with numerous studies confirming their efficacy against varied parasites^{14,15,19,20,23,29,33,37,39}. Roots and rhizomes are also recurrently tested^{24,57}, as are aerial parts^{27,41,58,59}. Less commonly, stem bark and fruits³² have been evaluated, still contributing valuable insights into plant-based anthelmintic development.

4.4 Negative Effects of Parasitic Diseases in Animals

The diversity of plant parts studied for anthelmintic activity reflects variations in bioactive compounds and the influence of local medicinal traditions^{54,55}, reinforcing the need for a holistic approach in evaluating medicinal plants. Animal parasitic diseases impact the economy, society, and public health⁶⁰. The reviewed studies address a wide spectrum of parasites—including helminths, protozoa, and others—affecting both humans and animals, with some species transmissible between them, such as *Ancylostoma duodenale*⁶³. Livestock parasites, including *Haemonchus contortus* in small ruminants⁶¹ and

Paramphistomum cervi in cattle⁶², cause direct economic losses, highlighting the urgency of developing new treatments.

4.5 Extraction Methods and Biological Tests

Most studies employed solvent-based extraction of bioactive compounds from different plant parts, using techniques such as ultrasound-assisted (UAE), microwave-assisted (MAE), pressurized liquid (PLE), and supercritical fluid extraction (SFE)⁶⁴. Anthelmintic activity was assessed *in vitro* and *in vivo*^{19,22,25,37,38,100} targeting survival, reproduction, and motility of parasites. Tests addressed different life stages—adult worms, larvae, and eggs—including adult worm motility inhibition⁵⁷, egg hatch inhibition¹⁷, and larval assays^{24,29}. Cytotoxicity evaluations on mammalian cells (e.g., mammary cells¹⁴, WI38 cells¹⁶) ensured safety. Some studies also examined activity against non-helminth microorganisms such as bacteria and fungi²⁷.

4.6 Safety and Efficacy of Medicinal Plants

Safety assessments included cell viability tests, while efficacy trials covered diverse organisms, such as *Haemonchus placei*¹⁴, *H. contortus* and *Trichostrongylus colubriformis*⁵⁸, *Eisenia foetida*⁵⁹, *Gyrodactylus kobayashi*²⁵, and *Marshallagia marshalli*¹³. Several plants and compounds showed strong anthelmintic potential: root extracts active against adult worms⁵⁷, ethyl acetate extracts with high efficacy⁵⁹, plumbagin²⁵, flavonoids and secoiridoid glycosides²⁷, and essential oils active against *M. marshalli*¹³. Others demonstrated limited or moderate efficacy, such as garlic, ineffective against equine strongyles³⁸, and MC010, moderately active against *Fasciola hepatica* but cytotoxic²³. These findings underscore both the promise and limitations of plant-derived anthelmintics.

4.7 Technological Prospecting: Patent Analysis

India, with a high prevalence of helminth infections such as *Ancylostoma duodenale*, faces increasing drug resistance, prompting the search for novel therapeutic agents^{65,66}. Approximately 300 helminth species can infect humans, particularly affecting children's health and quality of life. The patents analyzed investigated species including *Andrographis paniculata*, *Chenopodium album*, *Calotropis* sp., *Cocos nucifera*, *Operculina turpethum*, *Adhatoda vasica*, and *Momordica charantia*. While mostly of Asian origin, *C. nucifera* and *M. charantia* are also found in Brazil. *M. charantia* shows antimicrobial and anthelmintic

activity against *Fasciola hepatica* and *Strongyloides* spp.^{67,68}, while *A. paniculata*, rich in andrographolide, exhibits anti-inflammatory, antiviral, and antimalarial properties⁶⁹. *C. album* demonstrated efficacy against *Haemonchus contortus*, with molecular docking identifying isolariciresinol 4'-O- β -D-glucoside as a promising compound⁷⁰. *O. turpethum* roots contain lupel, betulin, β -sitosterol, and cycloartenol, all with antibacterial and antitumor activity^{71,72}. Two patents tested activity against *A. duodenale* and *Brugia malayi*.

Other studies employed experimental models. Hookworm, prevalent in areas lacking basic sanitation, is a significant cause of mortality, transmitted via the fecal–oral route or contact with contaminated soil, and may be asymptomatic or cause anemia depending on parasite load^{73,75}. *Brugia malayi* filariae, transmitted by insect bites, infect the human lymphatic system in their adult form, leading to elephantiasis—a condition with severe social stigma affecting mainly impoverished populations⁷⁵.

Mass drug administration remains a control strategy in high-incidence areas; however, it fosters resistance and is not fully effective⁷⁶. Four patents tested experimental parasite models (*Eisenia foetida*, *Pheretima posthuma*, and *Haemonchus contortus*), chosen due to challenges in cultivating pathogenic helminths, whose complex life cycles require costly logistics⁷⁷. Free-living, non-pathogenic helminths such as *E. foetida* and *P. posthuma* are commonly used because of their similarity to human intestinal parasites⁵⁹.

Extracts were the most common form tested (4 patents), followed by microencapsulation (1 patent) and ashes (1 patent). Concentrations ranged from 200 mg/mL (ethanolic extracts of *Adhatoda vasica* and *Momordica charantia*) to 62.5 μ g/mL (ethanolic and metallic extracts of *Calotropis* sp. and isolated compounds). All patents showed potential, with notable results for microencapsulated *Andrographis paniculata*, which reduced *A. duodenale* load at 5 mg, and ethanolic *Calotropis* sp., achieving 100% microfilaricidal activity at 125 μ g/mL. Patent filings spanned 2010–2022, peaking in 2020 with three registrations (Figure 1).

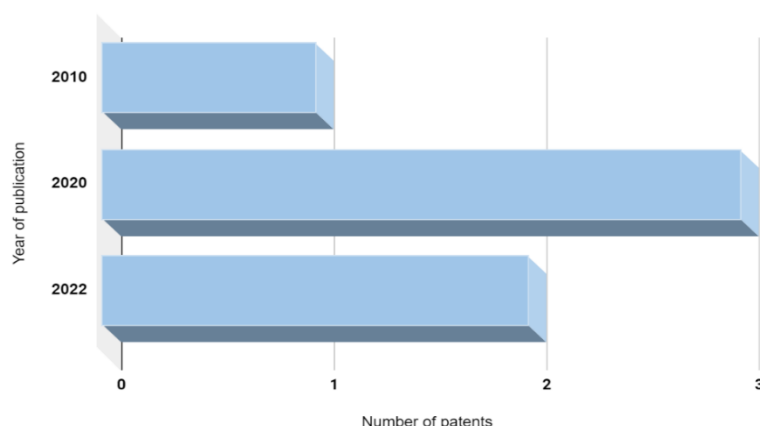


Figure 1 - Annual evolution of the number of patent registrations filed between 2010 and 2022.

The onset of the COVID-19 pandemic in 2020 stimulated global research and patenting activity, including innovations for emerging diseases⁷⁸. India, home to major producers of Active Pharmaceutical Ingredients (APIs) and vaccines, experienced a surge in health-related patents⁷⁹. The increase in patents for medicinal plant-based anthelmintic therapies in 2020 may reflect heightened global interest in natural medicines and alternative therapies during the pandemic, driven by greater health awareness⁸⁰. Investments in research, institutional–corporate–government collaborations, and the country’s rich plant diversity likely contributed to this growth^{81,82}.

Innovations in phytopharmaceuticals show notable advances over conventional treatments. An ethanolic extract exhibited microfilaricidal activity against *Brugia malayi* at low concentrations⁴², while a microencapsulated product demonstrated ovicidal and larvicidal effects against *Ancylostoma duodenale*, matching or surpassing positive controls⁴³. Plant formulations of *Chenopodium album* and *Operculina turpethum* effectively paralyzed and killed worms⁵⁹. Coconut shell ash proved active against *Pheretima posthuma* and *Haemonchus contortus*⁴⁶, and comparative assays of *Adhatoda vasica* and *Momordica charantia* yielded valuable insights for therapy development⁴⁷. These alternatives offer potential improvements in efficacy, accessibility, and public health outcomes.

The number of scientific articles exceeded patents in this field, reflecting the natural progression from research to commercialization. Academic studies often precede patent filings, focusing on exploratory and pre-clinical evaluations of safety and efficacy. Patent registration, being complex, costly, and time-intensive, further explains the lower filing volume despite active research.

5. CONCLUSION

Research into the therapeutic potential of medicinal plants against helminths has made significant progress, highlighting compounds such as plumbagin, flavonoids and essential oils as promising anthelmintic agents. The analysis of plant parts reveals a diversity of bioactive compounds, emphasizing the importance of a holistic approach in herbal research. The studies address a wide range of parasites, highlighting the need for new therapies to combat these pathogens. In vitro and in vivo methods are used to assess the efficacy and safety of compounds, reflecting a commitment to evidence-based pharmaceutical research. The increase in the number of patent filings in India in 2020 highlights the growing global interest in anthelmintic therapies based on medicinal plants and the investments in research and development in this area. These innovations reveal significant advances in phytopharmaceutical therapies for the treatment of parasitic conditions, offering promising alternatives to conventional treatments and potentially benefiting public health on a global scale.

REFERENCES

- 1 Adu-Gyasi D, Asante KP, Frempong MT, Gyasi DK, Iddrisu LF, Ankrah L, et al. Epidemiology of soil transmitted Helminth infections in the middle-belt of Ghana, Africa. *Parasite Epidemiol Control*. 2018;3:e00071. Disponível em: <https://doi.org/10.1016/j.parepi.2018.e00071>
- 2 Akinsanya B, Taiwo A, Adedamola M, C O. An investigation on the epidemiology and risk factors associated with soil-transmitted helminth infections in Ijebu East Local Government Area, Ogun State, Nigeria. *Sci Afr*. 2021;12:e00757. Disponível em: <https://doi.org/10.1016/j.sciaf.2021.e00757>
- 3 Mehraj V, Hatcher J, Akhtar S, Rafique G, Beg MA. Prevalence and factors associated with intestinal parasitic infection among children in an urban slum of Karachi. *PLoS One*. 2008;3:e3680. Disponível em: <https://doi.org/10.1371/journal.pone.0003680>
- 4 Madinga J, Polman K, Kanobana K, van Lieshout L, Brienen E, Praet N, et al. Epidemiology of polyparasitism with *Taenia solium*, schistosomes and soil-transmitted helminths in the co-endemic village of Malanga, Democratic Republic of Congo. *Acta Trop*. 2017;171:186–93. Disponível em: <https://doi.org/10.1016/j.actatropica.2017.03.019>
- 5 Chelkeba L, Mekonnen Z, Emanu D, Jimma W, Melaku T. Prevalence of soil-transmitted helminths infections among preschool and school-age children in Ethiopia: a systematic

review and meta-analysis. Glob Health Res Policy. 2022;7:21. Disponível em: <https://doi.org/10.1186/s41256-022-00239-1>

6 Daumerie D, Savioli L, Crompton DWT. Working to overcome the global impact of neglected tropical diseases: first WHO report on neglected tropical diseases. Geneva: World Health Organization; 2010. Disponível em: https://iris.who.int/bitstream/handle/10665/44440/9789241564090_eng.pdf

7 Tinkler SH. Preventive chemotherapy and anthelmintic resistance of soil-transmitted helminths – Can we learn nothing from veterinary medicine? One Health. 2020;9:100106. Disponível em: <https://doi.org/10.1016/j.onehlt.2019.100106>

8 Moyat M, Lebon L, Perdijk O, Wickramasinghe LC, Zaiss MM, Mosconi I, et al. Microbial regulation of intestinal motility provides resistance against helminth infection. Mucosal Immunol. 2022;15:1283–95. Disponível em: <https://doi.org/10.1038/s41385-022-00498-8>

9 Kuhn AGKN, Boeff DD, de Oliveira Carvalho L, Konrath EL. Ethnobotanical knowledge on native Brazilian medicinal plants traditionally used as anthelmintic agents – A review. Exp Parasitol. 2023;249:108531. Disponível em: <https://doi.org/10.1016/j.exppara.2023.108531>

10 Adak M, Kumar P. Herbal anthelmintic agents: a narrative review. J Tradit Chin Med. 2022;42:641–51. Disponível em: <https://doi.org/10.19852/j.cnki.jtcm.2022.04.007>

11 Silva FM de S e, Silva GC da, Carvalho RBF de, Nunes LCC. Topically Applied Anti-Inflammatory Properties of Medicinal Plants: A Systematic Review. Amazônia Sci Health. 2023;11(4):204–21. Disponível em: <https://doi.org/10.18606/2318-1419/amazonia.sci.health.v11n4p204-221>

12 Page MJ, McKenzie JE, Bossuyt PM, Boutron I, Hoffmann TC, Mulrow CD, et al. The PRISMA 2020 statement: an updated guideline for reporting systematic reviews. BMJ. 2021;372:n71. Disponível em: <https://doi.org/10.1136/BMJ.N71>

13 Ahmadzadeh Naghadeh Y, Malekifard F, Esmaeilnejad B. In vitro anthelmintic efficacy of medicinal plant essential oils against *Marshallagia marshalli*: evidence on oxidative/nitrosative stress biomarkers, DNA damage, and egg hatchability. Vet Parasitol. 2024;327:110138. Disponível em: <https://doi.org/10.1016/j.vetpar.2024.110138>

14 Ocampos GM, Ñumbay MT, Haddad M, Ambassa LMM, Alvarenga N, Hoste H. Two in vitro anthelmintic assays of four Paraguayan medicinal plants for proof of concept of the role of polyphenols in their biological activities and LC-HRMS analysis. J Ethnopharmacol. 2023;312:116453. Disponível em: <https://doi.org/10.1016/j.jep.2023.116453>

15 Toklo PM, Alowanou GG, Wouamba SCN, Assogba FM, Ahomadegbe MA, Sakirigui A, et al. UPLC-QToF-ESI-MS identification and anthelmintic activity of *Mitragyna inermis* (Willd.) Kuntze (Rubiaceae). Heliyon. 2023;9:e16448. Disponível em: <https://doi.org/10.1016/j.heliyon.2023.e16448>

16 Tchetan E, Ortiz S, Hughes K, Olounladé PA, Laurent P, Azando EVB, et al. HPLC-LTQ orbitrap mass spectrometry-based molecular networking for identifying anthelmintic

molecules in *Morinda lucida* Benth. S Afr J Bot. 2023;161:53–65. Disponível em: <https://doi.org/10.1016/j.sajb.2023.08.002>

17 Dicko A, Konaté A, Azokou A, Sylla Y, Tindano B, Ahoua ARC, et al. In vitro anthelmintic activity of *Euphorbia forskalii* J. Gay aqueous extracts evaluation on different life stages of *Haemonchus contortus*. Vet Parasitol Reg Stud Reports. 2023;44:100918. Disponível em: <https://doi.org/10.1016/j.vprsr.2023.100918>

18 Calvin BZ, Géorcelin AG, Phillipe BEK, Pascal OA, Laure MFA, Joyce KWJ, et al. The anti-parasitic effect of extract of *Ceiba pentandra* (L.) Gaertn is related to its anti-inflammatory, analgesic and anthelmintic activities on *Haemonchus contortus*. Clin Complement Med Pharmacol. 2023;3:100088. Disponível em: <https://doi.org/10.1016/j.ccmp.2023.100088>

19 Costa D de S, Leal CM, Cajas RA, Gazolla MC, Silva LM, Carvalho LSA de, et al. Antiparasitic properties of 4-nerolidylcatechol from *Pothomorphe umbellata* (L.) Miq. (Piperaceae) in vitro and in mice models with either prepatent or patent *Schistosoma mansoni* infections. J Ethnopharmacol. 2023;313:116607. Disponível em: <https://doi.org/10.1016/j.jep.2023.116607>

20 Idris OA, Wintola OA, Afolayan AJ. Anthelmintic potency of *Rumex crispus* L. extracts against *Caenorhabditis elegans* and non-targeted identification of the bioactive compounds. Saudi J Biol Sci. 2022;29:541–9. Disponível em: <https://doi.org/10.1016/j.sjbs.2021.09.026>

21 Kundu P, Debnath SL, Sadhu SK. Exploration of pharmacological and toxicological properties of aerial parts of *Blumea lacera*, a common weed in Bangladesh. Clin Complement Med Pharmacol. 2022;2:100038. Disponível em: <https://doi.org/10.1016/j.ccmp.2022.100038>

22 Queiroz MN, dos Santos Torres ZE, Pohlit AM, Ono EA, Affonso EG. Therapeutic potential of *Piper aduncum* leaf extract in the control of monogeneans in tambaqui (*Colossoma macropomum*). Aquaculture. 2022;552:738024. Disponível em: <https://doi.org/10.1016/j.aquaculture.2022.738024>

23 Chakroborty A, Pritchard D, Bouillon ME, Cervi A, Cookson A, Wild C, et al. Flukicidal effects of abietane diterpenoid derived analogues against the food borne pathogen *Fasciola hepatica*. Vet Parasitol. 2022;309:109766. Disponível em: <https://doi.org/10.1016/j.vetpar.2022.109766>

24 Kpabi I, Munsch T, Agban A, Théry-Koné I, Dorat J, Boudesocque-Delaye L, et al. *Cassia sieberiana* root bark used in traditional medicine in Togo: Anthelmintic property against *Haemonchus contortus* and tannins composition. S Afr J Bot. 2022;151:549–58. Disponível em: <https://doi.org/10.1016/j.sajb.2022.05.055>

25 Tu X, Duan C, Wu S, Fu S, Ye J. Identification of plumbagin as an effective chemotherapeutic agent for treatment of *Gyrodactylus* infections. Aquaculture. 2021;535:736372. Disponível em: <https://doi.org/10.1016/j.aquaculture.2021.736372>

- 26 Farhat F, Wasim S, Abidi SMA. Anthelmintic effect of thymoquinone against biliary amphistome, *Gigantocotyle explanatum*. Exp Parasitol. 2022;243:108421. Disponível em: <https://doi.org/10.1016/j.exppara.2022.108421>
- 27 Hussein D, El-Shiekh RA, Saber FR, Attia MM, Mousa MR, Atta AH, et al. Unravelling the anthelmintic bioactives from *Jasminum grandiflorum* L. subsp. *Floribundum* adopting in vitro biological assessment. J Ethnopharmacol. 2021;275:114083. Disponível em: <https://doi.org/10.1016/j.jep.2021.114083>
- 28 Ogedengbe-Olowofoyeku AN, Ademola IO, Wright CW, Idowu SO, Fatokun AA. Anthelmintic activity and non-cytotoxicity of phaeophorbide-a isolated from the leaf of *Spondias mombin* L. J Ethnopharmacol. 2021;280:114392. Disponível em: <https://doi.org/10.1016/j.jep.2021.114392>
- 29 Gogoi B, Tamuli KJ, Neipihoi, Bordoloi M, Sharma HK. Isolation and characterization of chemical constituents with in vitro anti-hypertensive and anthelmintic activities of *Cinnamomum bejolghota* (Buch.-Ham.) sweet leaves: An ethno medicinal plant of North East India. S Afr J Bot. 2021;140:161–6. Disponível em: <https://doi.org/10.1016/j.sajb.2021.04.004>
- 30 Minsakorn S, Watthanadirek A, Poolsawat N, Puttarak P, Chawengkirttikul R, Anuracpreeda P. The anthelmintic potentials of medicinal plant extracts and an isolated compound (rutin, C₂₇H₃₀O₁₆) from *Terminalia catappa* L. against *Gastrothylax crumenifer*. Vet Parasitol. 2021;291:109385. Disponível em: <https://doi.org/10.1016/j.vetpar.2021.109385>
- 31 Muhammad N, Ullah S, Rauf A, Atif M, Patel S, Israr M, et al. Evaluation of the anti-diarrheal effects of the whole plant extracts of *Cuscuta reflexa* Roxb in pigeons. Toxicol Rep. 2021;8:395–404. Disponível em: <https://doi.org/10.1016/j.toxrep.2021.02.013>
- 32 Wahab Obeng A, Boakye YD, Agana TA, Djameh GI, Boamah D, Adu F. Anti-trypanosomal and anthelmintic properties of ethanol and aqueous extracts of *Tetrapleura tetraptera* Taub. Vet Parasitol. 2021;294:109449. Disponível em: <https://doi.org/10.1016/j.vetpar.2021.109449>
- 33 Ramadwa TE, McGaw LJ, Adamu M, Madikizela B, Eloff JN. Anthelmintic, antimycobacterial, antifungal, larvicidal and cytotoxic activities of acetone leaf extracts, fractions and isolated compounds from *Ptaeroxylon obliquum* (Rutaceae). J Ethnopharmacol. 2021;280:114365. Disponível em: <https://doi.org/10.1016/j.jep.2021.114365>
- 34 Mongalo NI, Mashele SS, Makhafola TJ. *Ziziphus mucronata* Willd. (Rhamnaceae): its botany, toxicity, phytochemistry and pharmacological activities. Heliyon. 2020;6:e03708. Disponível em: <https://doi.org/10.1016/j.heliyon.2020.e03708>
- 35 Olmedo-Juárez A, Zarza-Albarran MA, Rojo-Rubio R, Zamilpa A, González-Cortazar M, Mondragón-Ancelmo J, et al. *Acacia farnesiana* pods (plant: Fabaceae) possesses anti-parasitic compounds against *Haemonchus contortus* in female lambs. Exp Parasitol. 2020;218:107980. Disponível em: <https://doi.org/10.1016/j.exppara.2020.107980>

- 36 Zhang Y, Tan X, Tu X, Ling F, Wang G. Efficacy and antiparasitic mechanism of curdione from *Curcuma zedoaria* against *Gyrodactylus kobayashii* in goldfish. *Aquaculture*. 2020;523:735186. Disponível em: <https://doi.org/10.1016/j.aquaculture.2020.735186>
- 37 Dkhil MA, Thagfan FA, Hassan A Moniem S, Al-Shaebi EM, Abdel-Gaber R, Al-Quraishy S. Anthelmintic, anticoccidial and antioxidant activity of *Salvadora persica* root extracts. *Saudi J Biol Sci*. 2019;26:1223–6. Disponível em: <https://doi.org/10.1016/j.sjbs.2019.02.006>
- 38 Buono F, Pacifico L, Piantedosi D, Sgroi G, Neola B, Roncoroni C, et al. Preliminary observations of the effect of garlic on egg shedding in horses naturally infected by intestinal strongyles. *J Equine Vet Sci*. 2019;72:79–83. Disponível em: <https://doi.org/10.1016/j.jevs.2018.10.025>
- 39 Banerjee T, Singh A, Kumar S, Dhanani T, Gajbhiye NA, Koley TK, et al. Ovicidal and larvicidal effects of extracts from leaves of *Andrographis paniculata* (Burm. f.) Wall. ex Nees against field isolates of human hookworm (*Ancylostoma duodenale*). *J Ethnopharmacol*. 2019;235:489–500. Disponível em: <https://doi.org/10.1016/j.jep.2019.02.021>
- 40 Feilcke R, Arnouk G, Raphane B, Richard K, Tietjen I, Andrae-Marobela K, et al. Biological activity and stability analyses of knipholone anthrone, a phenyl anthraquinone derivative isolated from *Kniphofia foliosa* Hochst. *J Pharm Biomed Anal*. 2019;174:277–85. Disponível em: <https://doi.org/10.1016/j.jpba.2019.05.065>
- 41 Cortes-Morales JA, Olmedo-Juárez A, Trejo-Tapia G, González-Cortazar M, Domínguez-Mendoza BE, Mendoza-de Gives P, et al. In vitro ovicidal activity of *Baccharis conferta* Kunth against *Haemonchus contortus*. *Exp Parasitol*. 2019;197:20–8. Disponível em: <https://doi.org/10.1016/j.exppara.2019.01.003>
- 42 Gallego FL, Avelino B da SS. Epidemiologia das doenças infecciosas parasitárias. *Rev FT*. 2023;1. Disponível em: <https://revistafatectq.com.br/index.php/revista/article/view/94>
- 43 Pedroso RDS, Andrade G, Pires RH. Plantas medicinais: uma abordagem sobre o uso seguro e racional. *Physis Rev Saúde Coletiva*. 2021;31:e310218. Disponível em: <https://doi.org/10.1590/S0103-73312021310218>
- 44 Borborema DC, Sousa HL, Vilar MS de A, Vilar D de A, Confessor MA. Plantas medicinais no tratamento de doenças parasitárias intestinais. *Anais II Congr Bras Ciênc Saúde*. 2017;3322:3222. Disponível em: <https://editorarealize.com.br/artigo/visualizar/29523>
- 45 Pires C de A, Mazola H, Jesus LF de, Mascarenhas LA, Viera LCDS. Interação planta medicamento e sua ação como antiparasitário: revisão integrativa. *Anais XIV Semin Estud Pesq Extensão FAMAM*. 2017;17:1–1. Disponível em: <https://unimam.com.br/wp-content/uploads/2020/05/interacao-planta-medicamento-e-sua-acao-como-antiparasitario-revisao-integrativa.pdf>
- 46 Soares SB, Moraes IL de, Caes AL. Etnobotânica e preservação cultural: tradição, comunidade, escola e educação ambiental. *Rev Cient Multidiscip Núcleo Conhecimento*.

- 2023:225–60. Disponível em:
<https://doi.org/10.32749/nucleodoconhecimento.com.br/educacao/etnobotanica>
- 47 Carneiro FM, Silva MJP da, Borges LL, Albernaz LC, Costa JDP. Trends of studies for medicinal plants in Brazil. Rev Sapiência Soc Saberes Prát Educ UEG Câmpus Iporá. 2014;3:44–75. Disponível em:
<https://www.revista.ueg.br/index.php/sapiencia/article/view/2954>
- 48 Ahmed H, Kilinc SG, Celik F, Kesik HK, Simsek S, Ahmad KS, et al. An inventory of anthelmintic plants across the globe. Pathogens. 2023;12:131. Disponível em:
<https://doi.org/10.3390/pathogens12010131>
- 49 Rajeswari VD. Anthelmintic activity of plants: a review. Res J Phytochem. 2014;8(3):57–63. Disponível em: <https://doi.org/10.3923/rjphyto.2014.57.63>
- 50 Spiegler V, Liebau E, Hensel A. Medicinal plant extracts and plant-derived polyphenols with anthelmintic activity against intestinal nematodes. Nat Prod Rep. 2017;34:627–43. Disponível em: <https://doi.org/10.1039/C6NP00126B>
- 51 Yang J, Chen WY, Fu Y, Yang T, Luo XD, Wang YH, et al. Medicinal and edible plants used by the Lhoba people in Medog County, Tibet, China. J Ethnopharmacol. 2020;249:112430. Disponível em: <https://doi.org/10.1016/j.jep.2019.112430>
- 52 Aderibigbe SA, Idowu SO, Olaniyi AA, Wright CW, Fatokun AA. Bioactivity and cytotoxicity profiling of vincosamide and strictosamide, anthelmintic epimers from *Sarcocephalus latifolius* (Smith) Bruce leaf. J Ethnopharmacol. 2021;265:113142. Disponível em: <https://doi.org/10.1016/j.jep.2020.113142>
- 53 Choudhary N, Khatik GL, Choudhary S, Singh G, Suttee A. In vitro anthelmintic activity of *Chenopodium album* and in-silico prediction of mechanistic role on *Eisenia foetida*. Heliyon. 2021;7:e05917. Disponível em: <https://doi.org/10.1016/j.heliyon.2021.e05917>
- 54 Zhang Y, Chen G, Zhou S, He L, Ayanniyi OO, Xu Q, et al. APDDD: Animal parasitic diseases and drugs database. Comp Immunol Microbiol Infect Dis. 2024;104:102096. Disponível em: <https://doi.org/10.1016/j.cimid.2023.102096>
- 55 Adduci I, Sajovitz F, Hinney B, Lichtmannsperger K, Joachim A, Wittek T, et al. Haemonchosis in sheep and goats, control strategies and development of vaccines against *Haemonchus contortus*. Animals. 2022;12:2339. Disponível em:
<https://doi.org/10.3390/ani12182339>
- 56 Ates C, Umur S. Paramphistome species in water buffaloes and intermediate hosts in the Kızılırmak Delta in Samsun Province, Turkey. Acta Parasitol. 2021;66:213–21. Disponível em: <https://doi.org/10.1007/s11686-020-00278-z>
- 57 Ngui R, Lim YAL, Traub R, Mahmud R, Mistam MS. Epidemiological and genetic data supporting the transmission of *Ancylostoma ceylanicum* among human and domestic animals. PLoS Negl Trop Dis. 2012;6:e1522. Disponível em:
<https://doi.org/10.1371/journal.pntd.0001522>

-
- 58 Barbero GF. Extraction and analysis of natural product in plant. *Agronomy*. 2021;11:415. Disponível em: <https://doi.org/10.3390/agronomy11030415>
- 59 Tuli R, Rawat AKS, Khatoon S, Srivastava S, Rastogi S, Pandey MM, et al. A phyto-pharmaceutical preparation useful for the treatment of filaria especially as a macrofilaricidal agent. Índia: IN2020202020; 2020.
- 60 Kumar S, Singh R, Banerjee T, Singh A. Microencapsulated herbal product with ovicidal and larvicidal properties against *Ancylostoma duodenale* (hookworm). Índia: IN202021057226; 2020.
- 61 Suttee A, Choudhary N. A novel formulation of *Chenopodium album* for anthelmintic activity. Índia: IN20201102376; 2020.
- 62 Suttee A, Choudhary N. A novel formulation of *Operculina turpethum* for anthelmintic activity. Índia: IN202011023772; 2020.
- 63 Gupta AK, Mishra AK, Singh R, Singh H, Singh MK, Mishra A, et al. *Cocos nucifera* (coconut) husk ash as anthelmintic drug substance. Índia: IN202211024662; 2022.
- 64 Nitalikar MM, Khade SY, Jadhav SD, Kute PB, Thombare AB, Gaikwad PP, et al. In-vitro comparative anthelmintic activity of *Adhatoda vasica* bark and *Momordica charantia* against *Pheretima posthuma*. Índia: IN202221032849; 2022.
- 65 Ajjampur SSR, Kaliappan SP, Halliday KE, Palanisamy G, Farzana J, Manuel M, et al. Epidemiology of soil transmitted helminths and risk analysis of hookworm infections in the community: results from the DeWorm3 Trial in southern India. *PLoS Negl Trop Dis*. 2021;15:e0009338. Disponível em: <https://doi.org/10.1371/journal.pntd.0009338>
- 66 Jayawardene KLT, Palombo EA, Boag PR. Natural products are a promising source for anthelmintic drug discovery. *Biomolecules*. 2021;11:1457. Disponível em: <https://doi.org/10.3390/biom11101457>
- 67 Laczkó-Zöld E, Bacsadi B, Horváth A, Csupor D. Development and validation of a RP-HPLC-DAD method for quantification of charantin in *Momordica charantia* products. *J Food Compos Anal*. 2021;104:104161. Disponível em: <https://doi.org/10.1016/j.jfca.2021.104161>
- 68 Poolperm S, Jiraungkoorskul W, Jiraungkoorskul W, Jiraungkoorskul W. An update review on the anthelmintic activity of bitter gourd, *Momordica charantia*. *Pharmacogn Rev*. 2017;11:31–4. Disponível em: https://doi.org/10.4103/phrev.phrev_52_16
- 69 Dai Y, Chen SR, Chai L, Zhao J, Wang Y, Wang Y. Overview of pharmacological activities of *Andrographis paniculata* and its major compound andrographolide. *Crit Rev Food Sci Nutr*. 2019;59(Suppl 1):S17–29. Disponível em: <https://doi.org/10.1080/10408398.2018.1501657>
- 70 Islam Z, Amin A, Paul GK, Hasan K, Rashid M, Saleh MA, et al. Anthelmintic, antioxidant, and cytotoxic activities of *Chenopodium album* against *Haemonchus contortus*:

a combined in vitro and in silico study. Inform Med Unlocked. 2023;37:101194. Disponível em: <https://doi.org/10.1016/j.imu.2023.101194>

71 Aggarwal BB, Prasad S, Reuter S, Kannappan R, Yadav VR, Park B, et al. Identification of novel anti-inflammatory agents from Ayurvedic medicine for prevention of chronic diseases: “Reverse pharmacology” and “Bedside to Bench” approach. Curr Drug Targets. 2011;12:1595–653. Disponível em: <https://doi.org/10.2174/138945011798109464>

72 Gupta S, Ved A. *Operculina turpethum* (Linn.) Silva Manso as a medicinal plant species: a review on bioactive components and pharmacological properties. Pharmacogn Rev. 2017;11:158. Disponível em: https://doi.org/10.4103/phrev.phrev_6_17

73 Furtado LFV, de Miranda RRC, Tennessen JA, Blouin MS, Rabelo ÉML. Molecular variability of the *Ancylostoma secreted Protein-2* (Aca-asp-2) gene from *Ancylostoma caninum* contributes to expand information on population genetic studies of hookworms. Exp Parasitol. 2023;253:108590. Disponível em: <https://doi.org/10.1016/j.exppara.2023.108590>

74 Ronquillo AC, Puelles LB, Espinoza LP, Sánchez VA, Luis Pinto Valdivia J. *Ancylostoma duodenale* as a cause of upper gastrointestinal bleeding: a case report. Braz J Infect Dis. 2019;23:471–3. Disponível em: <https://doi.org/10.1016/j.bjid.2019.09.002>

75 Servián A, Repetto SA, Lorena Zonta M, Navone GT. Human hookworms from Argentina: differential diagnosis of *Necator americanus* and *Ancylostoma duodenale* in endemic populations from Buenos Aires and Misiones. Rev Argent Microbiol. 2022;54:268–81. Disponível em: <https://doi.org/10.1016/j.ram.2022.05.005>

76 Casulli A. New global targets for NTDs in the WHO roadmap 2021–2030. PLoS Negl Trop Dis. 2021;15:e0009373. Disponível em: <https://doi.org/10.1371/journal.pntd.0009373>

77 Kamal M, Mukherjee S, Joshi B, Sindhu Z ud D, Wangchuk P, Haider S, et al. Model nematodes as a practical innovation to promote high throughput screening of natural products for anthelmintics discovery in South Asia: current challenges, proposed practical and conceptual solutions. Mol Biochem Parasitol. 2023;256:111594. Disponível em: <https://doi.org/10.1016/j.molbiopara.2023.111594>

78 Jemala M. Long-term research on technology innovation in the form of new technology patents. Int J Innov Stud. 2021;5:148–60. Disponível em: <https://doi.org/10.1016/j.ijis.2021.09.002>

79 George TK, Nair NP, Singh AK, Dilesh Kumar A, Roy AD, Mohan VN, et al. Development of a choice-framework for COVID vaccines in India using a multi-criteria decision analysis approach. Vaccine. 2023;41:3755–62. Disponível em: <https://doi.org/10.1016/j.vaccine.2023.04.062>

80 Divya M, Vijayakumar S, Chen J, Vaseeharan B, Durán-Lara EF. South Indian medicinal plants can combat deadly viruses along with COVID-19? – A review. Microb Pathog. 2020;148:104277. Disponível em: <https://doi.org/10.1016/j.micpath.2020.104277>